



Figure 1. Cornish-Bowden plot showing competitive inhibition of chorismate synthase by (6*S*)-6-fluoro-EPSP (**7c**). UV assays (275 nm) were carried out at 25 °C, pH 7.0 (triethanolamine hydrochloride, 50 mM), and included 1.2 millimoles of chorismate synthase, 20 μM NADPH, 10 μM FMN, 50 mM KCl, and 2.5 mM MgCl₂ in addition to substrate EPSP (**7a**) and inhibitor (6*S*)-6-fluoro-EPSP (**7c**) in a final volume of 1 mL. Inhibitor concentrations were 0, 0.5, 1, 2, 3.5, and 5 μM, and substrate concentrations were (■) 20 μM EPSP, (▲) 35 μM EPSP, and (○) 50 μM EPSP.

compound under conditions which would have easily detected a turnover rate 0.2% that of EPSP itself.¹⁷

Competition experiments were performed in which chorismate synthase was assayed at various fixed concentrations of EPSP in the presence of a range of concentrations of **7b** or **7c**. Figure 1 shows a Cornish-Bowden plot¹⁸ of the data obtained for (6*S*)-6-fluoro-EPSP (**7c**). The parallel plots clearly signify a competitive mode of inhibition. The inhibition constant K_i was determined from a Dixon plot.¹⁹ It is found that both fluoro-EPSPs show clean competitive inhibition with **7c** having an affinity an order of magnitude greater than **7b**: K_i ((6*S*)-6-fluoro-EPSP) = 0.2 ± 0.1 μM, K_i ((6*R*)-6-fluoro-EPSP) = 3.0 ± 0.3 μM. These values compare with K_i (iso-EPSP **3**) = 8.7 μM,⁷ and K_m (EPSP) = 2.2 μM.⁸ The lack of irreversible inhibition by either compound was confirmed by incubation of *N. crassa* chorismate synthase with 50 μM of each inhibitor at 25 °C. Over a period 1 h, no loss of enzyme activity was observed relative to a control which lacked inhibitor.

The lack of irreversible inhibition is inconsistent with a mechanism involving a covalent enzyme-intermediate adduct such as **4**. While the observation that both 6-fluoro-EPSPs are potent competitive inhibitors does not itself support or preclude any of the other mechanisms in Scheme I, it does provide a useful tool for future mechanistic studies of the enzyme.

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Supplementary Material Available: Preparation of **7b,c** from **5b,c**, spectroscopic characterization of **5b,c** and **7b,c**, and Dixon plot showing inhibition of chorismate synthase by **7c** (4 pages). Ordering information is given on any current masthead page.

(16) Chorismate formation is normally monitored by the appearance of the diene chromophore which has its λ_{max} at 275 nm. In these experiments, absorbance was monitored in the range 240–300 nm in order to accommodate a possible shift in the absorbance maximum due to a fluorine substituent.

(17) This experiment does not rigorously preclude either compound being a substrate but puts an upper limit on their turnover rate.

(18) Cornish-Bowden, A. *Biochem. J.* 1974, 137, 143–144.

(19) Dixon, M. *Biochem. J.* 1953, 55, 170–171.

Modulation of Physical and Chemical Properties of η -H₂ Complexes of Osmium Amines by Facile Substitution

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Since the discovery of the first dihydrogen complex by Kubas et al.,¹ many dihydrogen complexes have been synthesized. In 1971,² the preparation in our laboratory of [Os(en)₂H₂]²⁺ as the chloride salt was reported. It was described as a dihydride and was assigned a *cis* configuration on the strength of ¹H NMR results which revealed two sets of amine protons in equal number. Our investigation of the analogous species [Os(NH₃)₄H₂]²⁺, not heretofore reported, throws new light on that structural assignment and, as well, provides ready access to a series of complexes arrived at by the simple addition of a variety of ligands to these 16e⁻ moieties.

When [Os(NH₃)₄H₂](B(C₆H₅)₄)₂³ (**1**) is dissolved in (C-D₃)₂CO, the ¹H NMR spectrum reveals only two kinds of protons ascribable to the cation, in the abundance ratio 6:1 at δ = 3.82 ppm and -11.37 ppm, respectively. For the purposes of species differentiation, the value of T_1 for the coordinated hydrogen was also measured (T_1 = 572 ms, 20 °C), as it was for the other species to be dealt with. When a trace of acid, for example, HO₃SCF₃, is present, slow H/D exchange between the solvent and coordinated hydrogen ensues, and, in a partially exchanged sample, J_{HD} was measured as 4.0 Hz. When any of a large number of solutes is added in excess, among them acetonitrile (AN), pyridine (Py), imidazole (Im), I⁻, Cl⁻, D₂O, and Br⁻, δ (ppm) J_{HD} (Hz), and T_1 (ms, 400 MHz) change and new characteristic values are registered. (See Table I.) In every case except with D₂O and (C-D₃)₂CO as addend, the corresponding solid salt was also prepared,⁵ and dissolved, with no discernible differences in the ¹H NMR signals. Because the solute level is low (0.010 M), we can conclude, at least in the case of the labile systems I⁻, Br⁻, or Cl⁻ as addend, that in acetone the affinity of the osmium center for the ligand is very high. As expected, it is much reduced in D₂O as solvent.

Of potential anionic ligands, the only one among those we have introduced which does not change the values of δ , J_{HD} , and T_1 is PF₆⁻ (even CF₃SO₃⁻ produces a set of characteristic values). This indicates that neither B(C₆H₅)₄⁻ nor PF₆⁻ enters the coordination sphere of the osmium complex, a supposition which, in the case of the former at least, is reasonable and, in view of the bulk and almost spherical shape of PF₆⁻, is reasonable for it also. However, it leaves open the question of whether (CD₃)₂CO also is a ligand when the B(C₆H₅)₄⁻ and PF₆⁻ salts are dissolved. That this is in fact the case is indicated by observations made for [Os(en)₂H₂]²⁺, and it is therefore likely also the case for [Os(NH₃)₄H₂]²⁺.

In preparing [Os(en)₂H₂]²⁺, we followed the literature procedure,² but with the difference that, instead of Cl⁻, B(C₆H₅)₄⁻ was

(1) Kubas, G. J.; Ryan, R. R.; Swanson, B. I.; Vergamini, P. J.; Wasserman, J. J. *J. Am. Chem. Soc.* 1984, 106, 451–452.

(2) Malin, J.; Taube, H. *Inorg. Chem.* 1971, 10, 2403.

(3) [Os(NH₃)₄H₂](B(C₆H₅)₄)₂ was made by the following procedure: Os(NH₃)₄(O₃SCF₃)₃ (100 mg) in 15 mL of H₂O was reduced by Zn/Hg (3 g) for 3 h, and then 15 mL of 0.2 M NaB(C₆H₅)₄ solution was added. The resulting precipitate was dried under vacuum. Microanal. Calcd for [Os(NH₃)₄H₂](B(C₆H₅)₄)₂·2H₂O: C, 61.67; H, 6.25; N, 5.99. Found: C, 61.50; H, 6.20; N, 5.80. Yield: >70%.

(4) Li, Z.-W.; Harman, W. D.; Lay, P. A.; Taube, H. *Inorg. Chem.*, submitted.

(5) The preparation of the pyridine adduct is typical of the others. The compound [Os(NH₃)₄(H₂)·Py][B(C₆H₅)₄]₂ (**2**) was prepared by dissolving **1** (100 mg) in pyridine (5 mL); after 1 h, ether was added to cause precipitation. The precipitate was collected, washed with ether, and dried. Yield: 90%. Microanal. Calcd for [Os(NH₃)₄(H₂)·Py][B(C₆H₅)₄]₂·2H₂O: C, 62.81; H, 5.82; N, 6.91. Found: C, 62.76; H, 6.03; N, 6.64. ¹H NMR in (CD₃)₂CO (ppm): 8.83 (d, 2 H, Py), 8.14 (t, 1 H, Py), 7.75 (t, 2 H, Py), 7.40–6.70 (m, 40 H, C₆H₅), 3.74 (s, br, 12 H, 4 NH₃), -7.44 (s, 2 H, OsH₂).

Table I. ^1H NMR Data^a of Selected Dihydrogen Complexes $[\text{Os}^{\text{II}}(\text{NH}_3)_4(\text{H}_2)\text{L}]^{2+/1+}$

L	AN	Py	Im	I ⁻	Cl ⁻	D ₂ O	(CD ₃) ₂ CO	Br ⁻
$\delta(\text{H}_2)$, ppm	-7.80 ^c	-7.44	-7.43	-11.60	-10.78 ^c	-11.35	-11.37 ^{bc}	-11.29
J_{HD} , Hz	20.3	19.6	17.1	12.5	10.2	8.1	4.0	<2.5
T_1 , ms (20 °C)	221	148	131	142	1026	346	572	963
T_1 , ms (low temperature)	62 ^d	38 (min)	63 (min)	129 ^d	125 ^d	77 (min)	74 ^d	107 ^d

^a In (CD₃)₂CO as solvent, at 400 MHz; complexes at ca. 0.010 M level. ^b Identical values of parameters for PF₆⁻ and B(C₆H₅)₄⁻ salts. ^c No new peaks at -95 °C or above. ^d No minimum observed at -95 °C or above.

**Figure 1.** Trans-to-cis isomerization.

used as precipitant. Elemental analysis shows the composition of the solid to correspond to $[\text{Os}(\text{en})_2\text{H}_2\text{Cl}]\text{B}(\text{C}_6\text{H}_5)_4$.⁶ The solid was dissolved in acetone and treated with TlPF₆ to extract Cl⁻. After filtration, ether was added to the solution; the solid was collected, washed with ether, dried, and redissolved in (CD₃)₂CO, and the ^1H NMR spectrum was recorded. The proton signals for dihydrogen and for en prove to be different from those for the chloro complex.⁷ In addition, a peak is observed at $\delta = 2.39$ ppm (3.7 H atoms counted at the first measurement), which disappears ($t_{1/2} \sim 10$ min) without attendant alterations in the other peaks. We ascribe this behavior to the replacement of coordinated (CH₃)₂CO by (CD₃)₂CO.

We conclude that in solution each of the 16e⁻ species $[\text{Os}(\text{NH}_3)_4\text{H}_2]^{2+}$ and $[\text{Os}(\text{en})_2\text{H}_2]^{2+}$ adopts an additional ligand and that the nitrogen ligands are coplanar in both cases. In contrast to NH₃, the amine moieties in $[\text{Os}(\text{en})_2\text{H}_2]^{2+}$ are not free to rotate, and thus they respond to the different environments provided by the groups on the opposite faces of the molecular plane. X-ray crystallographic study of three compounds⁸ confirmed the trans, octahedral stereochemistry (H₂ being counted as one ligand). All π -acid ligands which we have introduced lead to values of J_{HD} in the high range, suggesting that such ligands cause the hydrogen atoms to draw together. The addition of a very strong π -acid such as C₆H₅CH₂NC leads to the immediate release of H₂. The product is a *trans*- $[\text{Os}(\text{en})_2]^{2+}$ complex, which shows ^1H NMR signals only for en. Presumably the axial positions are occupied by (CD₃)₂CO.

In every one of at least 10 different systems, where the complexes in acetone are kept at room temperature for many hours, the ^1H NMR signals change.⁹ For ammonia, the 12 protons appear in the ratio 2:1:1, the pattern expected for cis configurations (see Figure 1). For the bis(ethylenediamine) complexes in the cis configuration, a plane of symmetry is lacking even when the addend is a symmetrical ligand such as Cl⁻, and for the eight amine as well as the eight methylene protons, four to eight peaks appear, the number depending on the symmetry of the addend. In all cases

(6) Microanal. Calcd for $[\text{Os}(\text{en})_2(\text{H}_2)\text{Cl}]\text{B}(\text{C}_6\text{H}_5)_4$: C, 50.43; H, 5.69; N, 8.39; Cl, 5.32. Found: C, 50.18; H, 5.46; N, 8.22; Cl, 5.81. (en = H₂NCH₂CH₂NH₂). ^1H NMR (ppm, in (CD₃)₂CO): 5.31 (s, br, 4 H, NH₂), 4.03 (s, br, 4 H, NH₂), 2.67 (m, 4 H, -CH₂-), 2.11 (m, 4 H, -CH₂-), -12.57 (s, 2 H, OsH₂). $J_{\text{HD}} = 7.2$ Hz. T_1 (20 °C, 400 MHz) = 254 ms. T_1 (min) = 61 ms (400 MHz).

(7) ^1H NMR in (CD₃)₂CO: 5.75 (s, br, 4 H, NH₂), 4.34 (s, br, 4 H, NH₂), 2.76 (m, 4 H, -CH₂-), 2.34 (m, 4 H, -CH₂-), 2.39 (s, 3.7 H, (CH₃)₂CO), -13.19 (s, 2 H, OsH₂). T_1 (20 °C, 400 MHz) = 145 ms. T_1 (min) = 57 ms (400 MHz).

(8) X-ray crystal structure determinations on $[\text{Os}(\text{en})_2(\eta^2\text{-H}_2)\text{Cl}]\text{Cl}$, $[\text{Os}(\text{en})_2(\eta^2\text{-H}_2)\text{Br}]\text{Br}$, and $[\text{Os}(\text{en})_2(\eta^2\text{-H}_2)\text{py}](\text{PF}_6)_2\text{CH}_3\text{OH}$ by T. Hasegawa of this laboratory and H. Hope of the University of California, Davis, confirm the trans geometry in these cases. Though data were taken at low temperature, they are not refined enough to place the atoms of dihydrogen. Arrangements have been made with T. F. Koetzle of the Brookhaven National Laboratory for structure determination by neutron diffraction.

(9) For example, on leaving 2 in (CD₃)₂CO for 4 days, the original ^1H NMR peaks disappear and are replaced by a set of new peaks, which we assign as *cis*- $[\text{Os}(\text{NH}_3)_4(\text{H}_2)\text{Py}][\text{B}(\text{C}_6\text{H}_5)_4]$: 8.74 (d, 2 H, Py), 7.70 (t, 1 H, Py), 7.24 (t, 2 H, Py), 7.40-6.70 (m, 40 H, C₆H₅), 4.51 (s, br, 3 H, NH₃), 4.23 (s, br, 6 H, 2 NH₃), 3.58 (s, br, 3 H, NH₃), -7.32 (s, 2 H, OsH₂). $J_{\text{HD}} = 20.2$ Hz; T_1 (20 °C, 400 MHz) = 104 ms, T_1 (min) = 28 ms (400 MHz).

but one, conversion to the cis forms is more than 98% complete at equilibrium. The sole exception is $[\text{Os}(\text{NH}_3)_4(\text{H}_2)(\text{CH}_3\text{CN})]^{2+}$, where the conversion is only 90% complete.

The facile addition of both saturated and unsaturated ligands to the 16e⁻ moieties dealt with above provides a unique opportunity for a systematic study of the variation of the chemical and physical properties of η^2 complexes with composition and, trans and cis forms being available, also with geometry. Moreover, the use of the moieties as recognition probes, particularly for biological molecules, also suggests itself. Studies along both lines are in progress.

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The Structures of Quartromicins A₁, A₂, and A₃: Novel Macrocyclic Antiviral Antibiotics Possessing Four Tetrionic Acid Moieties

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In the course of a search for new antibiotics, novel antiviral antibiotics have been isolated from *Amycolatopsis orientalis* No. Q427-8. Non-ionic resin extraction of the fermentation broth (9.8 L) followed by chromatographic purification resulted in isolation of three active components, quartromicins A₁ (1; 57 mg), A₂ (2; 87 mg), and A₃ (3; 70 mg),¹ which contain a unique macrocyclic ring possessing four tetrionic acid moieties.

Quartromicin A₁ (1) [C₇₈H₈₈O₃₀, negative FABMS m/z 1525 (M - 2 + Na)⁻, 1541 (M - 2 + K)⁻] and quartromicin A₃ (3) [C₇₈H₉₂O₃₀, m/z 1545 (M - 2 + K)⁻] were isolated as colorless amorphous salts.² The ¹³C NMR spectra of 1 and 3 exhibit 39 well-defined carbon signals, and therefore, both antibiotics must possess symmetrical dimeric structures.

The ¹H and ¹³C NMR spectra³ of 1, 2, and 3 suggest the presence of a sugar, which was identified as D-galactose after isolation from the acid hydrolysis (MeOH/HCl) products. Although the aglycons, degalactosylquartromicins A₁-A₃,⁴ were isolated, the yield was rather poor (ca. 50%) and the aglycons were poorly soluble in NMR solvents. Because 3 could be purified most easily, structure determination was performed largely on the salt of 3. (Structure elucidation will be interpreted on the monomeric fragments hereafter.)

Routine 2D NMR³ analyses led to partial structures 4 and 5, which accounted for 33 of 39 carbons in the monomeric fragment of 3, but the following spectral properties, which were assigned

(1) Tsunakawa, M.; Tenmyo, O.; Tomita, K.; Naruse, N.; Kotake, C.; Miyaki, T.; Konishi, M.; Oki, T. *J. Antibiot.*, submitted for publication.

(2) The metal composition of the salt (mol %) of 3 was analyzed as follows: Na, 70%; K, 19%; Ca, 10%; Mg, 1%.

(3) The 1D and 2D NMR spectra are available as supplementary material.

(4) Degalactosyl-1: C₆₆H₆₈O₂₀, negative FABMS m/z 1201 (M - 2 + Na)⁻, 1217 (M - 2 + K)⁻.